

SCN5A mutations in 442 neonates and children: genotype–phenotype correlation and identification of higher-risk subgroups

Alban-Elouen Baruteau^{12,3,4*}, Florence Kyndt⁴ Elijah R. Behr¹ Arja S. Vink^{5,6}, Matthias Lachaud⁴ Anna Joong⁷ Jean-Jacques Schott⁴ Minoru Horie⁸ Isabelle Denjoy⁹ Lia Crotti¹⁰ Wataru Shimizu^{11,12}, Johan M. Bos^{13,14,15}, Elizabeth A. Stephenson¹⁶ Leonie Wong¹ Dominic J. Abrams¹⁷ Andrew M. Davis^{18,19}, Annika Winbo^{20,21}, Anne M. Dubin²² Shubhayan Sanatan²³ Leonardo Liberman⁷ Juan Pablo Kaski^{24,25}, Boris Rudic^{26,27}, Sit Yee Kwok²⁸ Claudine Rieubland²⁹ Jacob Tfelt-Hansen^{30,31}, George F. Van Hare³² Béatrice Guyomarc'h-Delasalle⁴ Nico A. Blom⁵ Yanushi D. Wijeyeratne¹ Jean-Baptiste Gourraud⁴ Hervé Le Marec⁴ Junichi Ozawa⁸ Véronique Fressart³³ Jean-Marc Lupoglazoff²⁴ Federica Dagradi⁴⁰ Carla Spazzolini⁴⁰ Takeshi Aiba¹¹ David J. Tester^{13,14,15}, Laura A. Zahavich¹⁶ Virginie Beauséjour-Ladouceur¹⁷ Mangesh Jadhav¹⁸ Jonathan R. Skinner^{20,21}, Sonia Franciosi²³ Andrew D. Krahn²³ Mena Abdelsayed³⁵ Peter C. Ruben³⁵ Tak-Cheung Yung²⁸ Michael J. Ackerman^{13,14,15}, Arthur A. Wilde^{6,36}, Peter J. Schwartz¹⁰ and Vincent Probst⁴

¹Cardiology Clinical Academic Group, Molecular and Clinical Sciences Research Institute, St George's University of London, London, UK; ²Department of Congenital Cardiology, Evelina London Children's Hospital, Guy's and St Thomas' NHS Foundation Trust, London, UK; ³M3C CHU de Nantes, Fédération des Cardiopathies Congénitales, Nantes, F-44000, France; ⁴L'institut du thorax, INSERM, CNRS, UNIV Nantes, CHU Nantes, Nantes, France; ⁵Department of Pediatric Cardiology, Academic Medical Center, Amsterdam, The Netherlands; ⁶Department of Clinical and Experimental Cardiology, Heart Centre, Academic Medical Center, Amsterdam, The Netherlands; ⁷Division of Pediatric Cardiology, Morgan Stanley Children's Hospital, New York Presbyterian Hospital, Columbia University Medical Center, New York, NY, USA; ⁸Department of Cardiovascular and Respiratory Medicine, Shiga University of Medical Sciences, Otsu, Japan; ⁹AP-HP, Hôpital Bichat, Service de Cardiologie, Université Denis Diderot, Paris, France; ¹⁰Center for Cardiac Arrhythmias of Genetic Origin, IRCCS Istituto Auxologico Italiano, Milano, Italy; ¹¹Department of Cardiovascular Medicine, National Cerebral and Cardiovascular Center, Suita, Osaka, Japan; ¹²Department of Cardiovascular Medicine, Nippon Medical School, Tokyo, Japan; ¹³Division of Heart Rhythm Services, Department of Cardiovascular Diseases, Mayo Clinic, Rochester, MN, USA; ¹⁴Division of Pediatric Cardiology, Department of Pediatrics, Mayo Clinic, Rochester, MN, USA; ¹⁵Department of Molecular Pharmacology & Experimental Therapeutics, Windland Smith Rice Sudden Death Genomics Laboratory, Mayo Clinic, Rochester, MN, USA; ¹⁶The Hospital for Sick Children, Labbatt Family, Heart Centre, University of Toronto, Toronto, Canada; ¹⁷Inherited Cardiac Arrhythmia Program, Boston Children's Hospital, Harvard Medical School, Boston, MA, USA; ¹⁸Department of Cardiology, The Royal Children's Hospital, Melbourne, Australia; ¹⁹Murdoch Children's Research Institute and University of Melbourne, Melbourne, Australia; ²⁰Greenlane Paediatric and Congenital Cardiac Services, Starship Children's Hospital, Auckland, New Zealand; ²¹Department of Paediatrics, Child and Youth Health, University of Auckland, Auckland, New Zealand; ²²Division of Pediatric Electrophysiology, Lucile Packard Children's Hospital, Stanford University, Palo Alto, CA, USA; ²³Divisions of Cardiology, Department of Pediatrics and Medicine, British Columbia Children's Hospital, University of British Columbia, Vancouver, BC, Canada; ²⁴Department of Cardiology, Centre for Inherited Cardiovascular Diseases, Great Ormond Street Hospital for Children, London, UK; ²⁵Institute of Cardiovascular Science, University College London, London, UK; ²⁶Medical Faculty Mannheim of the University of Heidelberg, 1st Department of Medicine, Mannheim, Germany; ²⁷DZHK (German Centre for Cardiovascular Research), Mannheim, Germany; ²⁸Department of Paediatric Cardiology, Queen Mary Hospital, The University of Hong Kong, Hong Kong SAR, China; ²⁹Division of Human Genetics, Department of Pediatrics, Inselspital, University of Bern, Switzerland; ³⁰Faculty of Health and Medical Science, Department of Clinical Medicine, University of Copenhagen, Copenhagen, Denmark; ³¹Department of Forensic Medicine, Faculty of Medical Sciences, University of Copenhagen, Denmark; ³²Division of Cardiology, Department of Pediatrics, Washington University in St. Louis School of Medicine, St. Louis, MO, USA; ³³AP-HP, Hôpital Pitié Salpêtrière, Service de Biologie Moléculaire, Paris, France; ³⁴AP-HP, Hôpital Robert Debré, Service de Cardiologie Pédiatrique, Paris, France; ³⁵Department of Biomedical Physiology and Kinesiology, Simon Fraser University, Burnaby, Canada; and ³⁶Princess Al-Jawhara Al-Brahim Centre of Excellence in Research of Hereditary Disorders, Jeddah, Kingdom of Saudi Arabia

Received 23 October 2017; revised 30 April 2018; editorial decision 7 June 2018; accepted 1 July 2018; online publish-ahead-of-print 27 July 2018

* Corresponding author. Tel:+442087252994, Fax:+442087253416, Email: alban-elouen.baruteau@u-bordeaux.fr

Published on behalf of the European Society of Cardiology. All rights reserved. © The Author(s) 2018. For permissions, please email: journals.permissions@oup.com.

Aims

To clarify the clinical characteristics and outcomes of children with *SCN5A*-mediated disease and to improve their risk stratification.

Methods and results

A multicentre, international, retrospective cohort study was conducted in 25 tertiary hospitals in 13 countries between 1990 and 2015. All patients ≤ 16 years of age diagnosed with a genetically confirmed *SCN5A* mutation were included in the analysis. There was no restriction made based on their clinical diagnosis. A total of 442 children {55.7% boys, 40.3% probands, median age: 8.0 [interquartile range (IQR) 9.5] years} from 350 families were included; 67.9% were asymptomatic at diagnosis. Four main phenotypes were identified: isolated progressive cardiac conduction disorders (25.6%), overlap phenotype (15.6%), isolated long QT syndrome type 3 (10.6%), and isolated Brugada syndrome type 1 (1.8%); 44.3% had a negative electrocardiogram phenotype. During a median follow-up of 5.9 (IQR 5.9) years, 272 cardiac events (CEs) occurred in 139 (31.5%) patients. Patients whose mutation localized in the C-terminus had a lower risk. Compound genotype, both gain- and loss-of-function *SCN5A* mutation, age ≤ 1 year at diagnosis in probands and age ≤ 1 year at diagnosis in non-probands were independent predictors of CE.

Conclusion

In this large paediatric cohort of *SCN5A* mutation-positive subjects, cardiac conduction disorders were the most prevalent phenotype; CEs occurred in about one-third of genotype-positive children, and several independent risk factors were identified, including age ≤ 1 year at diagnosis, compound mutation, and mutation with both gain- and loss-of-function.

Keywords

Brugada syndrome • Genotype–phenotype correlation • Long QT syndrome • Progressive cardiac conduction disorders • *SCN5A* • Sodium channelopathy

Introduction

Mutations in the gene (*SCN5A*) encoding the alpha subunit of the cardiac sodium channel (NaV1.5) cause type 3 long QT syndrome (LQT3),¹ type 1 Brugada syndrome (BrS-1),^{2,3} progressive cardiac conduction disorders (PCCD),^{3,4} atrial standstill and sick sinus syndrome (SSS),⁵ familial atrial fibrillation (AF),⁶ multifocal ectopic Purkinje-related premature contractions (MEPPC),⁷ dilated cardiomyopathy (DCM),⁸ and sudden infant death syndrome (SIDS).^{9,10} Some patients with *SCN5A* mutations are predisposed to sudden cardiac death (SCD), independently of age. A cardiac sodium channelopathy comprises a substantial proportion of aborted cardiac arrest (ACA) in children and adolescents.¹¹ Cardiac sodium channelopathies are diagnosed in infancy and early childhood following symptoms, sudden death, or family screening.^{12,13} Due to cascade genetic screening, the number of detected asymptomatic children with a *SCN5A* mutation is increasing. There is a significant variation in management of these asymptomatic *SCN5A* mutation-positive children amongst paediatric electrophysiologists.¹⁴ This is due to their relative rarity in the paediatric population. Therefore, challenging questions in clinical practice remain unanswered, and risk stratification is inadequate. This study aimed to assess the genotype–phenotype relationship and the risk analysis of cardiac sodium channelopathies in a large cohort of infants and children in order to improve their management.

Methods

Study design

A multicentre, international, retrospective cohort study was conducted in 25 tertiary hospitals in 13 different countries from January 1990 to December 2015. Institutional review board approval was obtained from

all participating institutions. All deceased and living patients ≤ 16 years of age diagnosed with a genetically confirmed *SCN5A* mutation were eligible for the study. There were no restrictions to the clinical diagnoses. Patients without a baseline electrocardiogram (ECG) were excluded from the analysis.

Clinical investigations

In all patients, demographic data, personal and family history (FH), mode of presentation, ECGs, echocardiography, treatment, and major cardiac events (MCEs) throughout follow-up were ascertained. Electrolyte and metabolic disturbances were excluded through laboratory tests. Study physicians gave their patients information about lifestyle modifications, such as aggressive antipyretic measures, the need for ECG monitoring during fever episodes, and avoidance of appropriate proarrhythmic drugs. Therapeutic management of the patients was based on the clinical judgement of the referring cardiologist. In case of device implantation, pacemaker (PM) type and mode of pacing, or implantable cardioverter-defibrillator (ICD) type and number of appropriate/inappropriate shocks were noted, as well as other device-related complications.

Genetic analysis

Mutation analysis of the *SCN5A* gene followed standard accepted protocols for genetic testing. Amino acid numbering was made according to transcription variant 1 of *SCN5A* (http://www.ncbi.nlm.nih.gov/NM_198056) and the predicted structure reported by Wang *et al.*,¹⁵ according to which the NaV1.5 alpha subunit protein consists of four transmembrane domains, each composed of six segments. The biophysical properties, type, and topological location of *SCN5A* mutations were determined on the basis of previously published data.^{16,17} All variants were reclassified by a group of authors (A.E.B., F.K., E.R.B., V.P.) at the time of this analysis according to the recommendation of the American College of Medical Genetics.¹⁸ *SCN5A* variants with minor allele frequency $>0.1\%$ in ExAC database (Exome Aggregation Consortium, Cambridge, MA, USA) and neutral synonymous variants were excluded.

Variants were then classified into three groups: missense pathogenic; non-missense pathogenic including truncating variants (non-sense, splice acceptor, splice donor, and frameshift mutations) and in-frame indels; and variants of uncertain significance (VUS). Missense variants were classified as pathogenic/likely pathogenic or VUS using generally accepted criteria¹⁸: disease-causative mutation databases, localization to highly conserved amino acid residues/key functional domains, co-segregation of the variant with the disease phenotype, evidence of perturbed ion channel function through *in vitro* functional studies. In case of double *SCN5A* mutation, patients were considered for risk analysis according to mutation location only if both mutations had the same location.

Statistical analysis

Continuous data were presented as mean (\pm standard deviation) or median [interquartile range (IQR)] based on the distribution. Categorical variables were presented as counts (proportions). The Mann–Whitney *U* and Kruskal–Wallis tests were performed to test for statistical differences in continuous parameters between two or more groups, respectively. The χ^2 or the Fisher exact test (based on expected frequency) were used to compare categorical variables between groups. Bonferroni method was used for *post hoc* tests. We adjusted *P*-value level on number of hypothesis tested. The Kaplan–Meier method estimator was used to assess the time to a first MCE. A Cox proportional-hazards regression analysis with random effect on family [with hazard ratios (HR) and confidence intervals (CI)] was used to evaluate the independent risk of clinical- and genetic-factors of interest for first MCE. From univariate analysis, we selected variables with *P*-value <0.10 (statistical criterion) and looked at multicollinearity between variables. For the multivariate model, we kept the following variables: proband, age <1 year at diagnosis, phenotype at baseline, genotype, location, HR, atrioventricular block (AVB), RBBB, and supraventricular arrhythmia. Variables were eliminated from highest to lowest *P*-values but remained in the final model if the *P*-value was less than 0.05 or seem to be confounders (more than 10% change in estimate). Final multivariable Cox model was stratified by phenotype (LQT3, PCCD, overlap phenotype, and ECG phenotype-negative) at baseline to relax the assumption of proportional hazards. All two-way interactions between pairs of predictors in the model were tested, one at a time. The mean event rate per year was evaluated by the number of events occurring during the follow-up divided by the number of patients multiplied by the average duration of follow-up. A *P*-value <0.05 was considered statistically significant when no Bonferroni correction was made. All *P*-values are two-sided. Due to the small number of patients in BrS-1, DCM and SSS phenotypes, these were not included in all the analysis. Data were analysed with the SAS packages (SAS Institute Inc. version 9, 4, Cary, NC, USA).

Results

A total of 442 children [246 boys (56%), 178 probands (40%), and median age at diagnosis of 8.0 (IQR 9.5) years] from 350 distinct families were eligible for the study.

Baseline clinical characteristics

Most of the patients (68%) were asymptomatic at diagnosis (Supplementary material online, Figure S1). The four ‘major’ ECG phenotypes at baseline were isolated PCCD (26%), overlap phenotype (16%), isolated LQT3 (11%), and isolated BrS1 (2%); 196 (44%) patients had a negative ECG phenotype at baseline (Figure 1). Clinical characteristics of each patients’ group are detailed in Supplementary

material online. All groups had similar gender distribution ($P=0.13$) and median age at diagnosis ($P=0.32$). The proportion of probands differed among groups ($P=0.02$). The mode of presentation also differed ($P<0.001$), an initial cardiac arrest being more frequent in overlap phenotype patients [16/69 (23%), $P=0.0001$], isolated PCCD patients [20/113 (18%), $P=0.002$], and isolated LQT3 patients [11/47 (23%), $P=0.0005$] compared to negative ECG phenotype patients [13/196 (7%)] (Supplementary material online, Table S1).

Clinical outcomes

Overall there were 272 MCEs in 139 (31%) patients during a median follow-up period of 5.9 years (IQR 5.9). Fifty (11%) patients had recurrent MCEs on treatment. Of the 77 (17%) ICD-implanted patients, 100 appropriate shocks were delivered in 28 (36%) patients during a median follow-up period of 3.3 years (Supplementary material online, Table S2). Inappropriate ICD shocks occurred in nine patients (12%; T wave oversensing in seven patients, AF in one, and lead fracture in one). The four ‘major’ ECG phenotypes at baseline developed as follows:

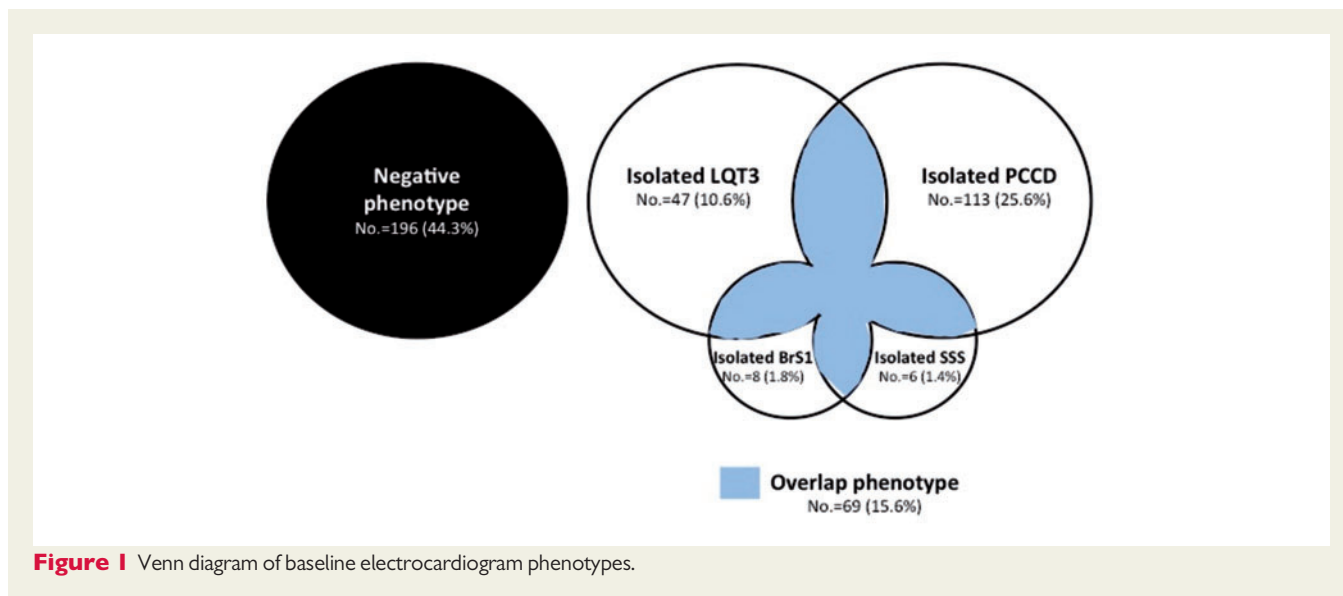
Isolated progressive cardiac conduction disorder patients

At a median follow-up of 5.7 (0.0–35.7) years, 26/113 (23%) patients kept an isolated PCCD phenotype; 13/113 (11%) had received PM implantation at a median age of 5.42 (0.06–15.58) years; 85% of PCCD patients had their first PM insertion by the age of 11; permanent PM were implanted for symptomatic bradycardia in 7/13 patients (syncope in 5, exercise-induced dyspnoea in 2), whilst the indications were prophylactic in 6/13 patients, including a mean daytime heart rate <50 b.p.m. in 4 children >1 year of age and ventricular pauses longer than 3 RR intervals in 2; 38/113 (34%) experienced ≥ 1 MCE, the first of which being cardiac arrest [18% including 3 documented ventricular tachycardia (VT), 1 polymorphic VT with torsades de pointes (TdP), and 1 ventricular fibrillation (VF)], SIDS (2%), or syncope (14%). At the time of their event, PCCD patients presented with the association of an AVB and right bundle branch block (RBBB) (17/38, 45%), an isolated first-degree AVB (13/38, 34%), an isolated complete RBBB (4/38, 10.5%) or a trifascicular block (4/38, 10.5%).

Two patients died (one during infancy, one SCD) and one required heart transplantation for intractable arrhythmias; although none of them underwent a sodium-channel blocker challenge, all three patients maintained an isolated cardiac conduction disorders phenotype throughout follow-up.

Overlap phenotype patients

After 5.7 (0.0–45.7) years, 34/69 (50%) patients had pharmacological treatment (beta-blocker: 39%, sodium channel blocker: 22% according to the combination of phenotypes, see Supplementary material online, Table S3); PM or ICD had been implanted in 10/69 (14%) and 17/69 (25%) respectively. At least one MCE occurred in 31/69 patients (45%; 1 recurrence in 6 patients, 2 recurrences in 1 patient, and ≥ 2 recurrences in 5 patients). Three patients died from SCD and one required extracorporeal membrane oxygenation support and was then transplanted for intractable arrhythmias.



Isolated type 3 long QT syndrome patients

At a median follow-up of 5.9 (0.0–26.5) years, 32/47 (68%) patients received a beta-blocker, coprescribed with a sodium channel blocker in 10 (21%), 3 (6%) had undergone left cardiac sympathetic denervation and PM and ICD implantation occurred in 3 (6%) and 11 (23%), respectively.

Major cardiac event occurred in 25 patients [53%, 5/25 (11%) \leq 1 year of age, 1/25 (4%) on beta-blocker at the time of the event] (Supplementary material online, Table S4). The first MCE was a SCD (2/47: 4%, including 1 during infancy), an ACA (19%) or a syncope (30%). Nine patients experienced more than one MCE. At the time of the first recurrent event, 7/9 patients were receiving beta-blocker therapy (Supplementary material online, Table S5); three patients experienced several recurrences under a coprescription of beta-blocker and mexiletine. Seven ICD shocks (6 appropriate, 1 inappropriate) were delivered in 3/11 (27%) implanted patients. Six (13%) patients died throughout follow-up, three of them had experienced a MCE in the first year of life.

Isolated type 1 Brugada syndrome patients

After 8.1 (1.8–15.7) years, 3/8 (37%) symptomatic BrS1 patients had an ICD (2.8, 11.5, and 18.8 years at implantation). They had presented with syncope (2 patients) or documented VT. One of them experienced a fever-associated VF-induced appropriate ICD shock at 13 years whilst under treatment. No death occurred. The five remaining patients were asymptomatic and left untreated.

Negative electrocardiogram phenotype patients

One hundred and ninety-six patients [44%, 52% boys, 33% probands, median age at diagnosis: 8.8 (IQR 8.7) years] had a normal ECG at baseline and underwent genetic screening because of cardiac arrest (7%), syncope (13%), or because of familial screening in asymptomatic patients (80%). A FH of either SCD/ICD implantation or PCCD/PM implantation was noted in 55% and 15%, respectively.

Of the 196 phenotype-negative patients, 27% developed an ECG phenotype throughout follow-up [5.9 (0.4–26.5) years], represented by an isolated PCCD phenotype (13%), an isolated LQT3 (5%), an isolated BrS1 (5%), or an overlap phenotype (4%), whereas 73% remained phenotype-negative. At least one MCE occurred in 40 (20%).

Of the 39 (20%) symptomatic, negative ECG phenotype patients, 26 received a beta-blocker. All but one negative ECG phenotype patients who experienced MCEs during follow-up were already symptomatic at diagnosis. Twelve experienced at least one recurrent MCE at a median delay of 3.9 (9.6) years since the diagnosis [median age of recurrent event: 3.0 (4.3) years]. All but one were treated by beta-blocker therapy at the time of the recurrent MCE; Of these 12 children, 8 kept a negative ECG phenotype at last visit, whereas 4 were further diagnosed with an isolated LQT3 phenotype and, despite additional treatment with mexiletine, experienced further recurrent MCEs leading to left cardiac sympathetic denervation (LCSD) and ICD implantation.

The vast majority (156/157, 99%) of the asymptomatic, negative ECG phenotype children remained asymptomatic throughout follow-up; one patient (0.6%) however became later symptomatic: this was a 5 year-old female patient with a normal ECG at familial screening; she was further diagnosed with an isolated LQT3 on follow-up ECGs at age 13 (QTc 491 ms) and received mexiletine; at age 18, she presented with an electrical storm whilst receiving mexiletine (500 mg morning, 250 mg afternoon, and 500 mg evening), leading to ICD implantation.

Genetic characteristics

The 442 *SCN5A* genotype-positive children had 185 independent *SCN5A* variants (Supplementary material online, Table S5). Three (0.7%) patients harboured a double heterozygous *SCN5A* mutation; 9 (2%) had a compound genotype with an additional disease-causing mutation in another gene: *KCNQ1* (3 patients), *KCNH2* (4 patients), *RYR2* (1 patient), or *CACNA1C* (1 patient). A loss-of-function mutation was found in 178 (40%) patients whereas, 87 (20%) had a gain-of-function mutation, 85 (19%) a both gain- and loss-of-function

Table 1 Risk analysis for major cardiac event (n = 442)

	No MCE (n = 303)	MCE (n = 139)	Analysis	HR (95% CI)	P-value
Clinical characteristics					
Male, n (%)	169 (55.8)	77 (55.4)	Yes vs. No	1 (0.7–1.5)	0.87
Proband, n (%)	75 (24.8)	103 (74.1)	Yes vs. No	7.8 (5.1–12.1)	<0.0001
Age ≤1 year at diagnosis, n (%)	34 (11.2)	41 (29.5)	Yes vs. No	11.3 (6.7–18.9)	<0.0001
Baseline ECG phenotype					
Isolated LQT3, n (%)	22 (7.3)	25 (18.0)	Yes vs. No	1.9 (1.1–3.1)	0.01
Isolated BrS-1, n (%)	5 (2.0)	3 (2.2)	Yes vs. No	1.2 (0.3–4.4)	0.69
Isolated PCCD, n (%)	75 (24.7)	38 (27.3)	Yes vs. No	1.2 (0.8–1.8)	0.29
Isolated DCM, n (%)	3 (0.9)	0 (0.0)	Yes vs. No	Not applicable	0.32 ^a
Isolated SSS, n (%)	4 (1.3)	2 (1.4)	Yes vs. No	0.9 (0.2–4.3)	0.84
Overlap phenotype, n (%)	38 (12.5)	31 (22.3)	Yes vs. No	1.9 (1.2–3.1)	0.004
Negative ECG phenotype, n (%)	156 (51.5)	40 (28.8)	Yes vs. No	0.4 (0.3–0.6)	<0.001
First available ECG characteristics ^a					
Median age at ECG (years) (IQR)	8.2 (8.4)	7.6 (12.8)	Unit = 2	0.8 (0.7–0.9)	<0.0001
Heart rate, b.p.m. (IQR)	79 (26.7)	77 (47.1)	Unit = 20	1.1 (1.0–1.3)	0.005
PR interval, ms (IQR)	160 (42)	160 (41)	Unit = 20	1.0 (0.9–1.1)	0.52
QRS complex, ms (IQR)	80 (24)	80 (40)	Unit = 20	1.0 (0.8–1.2)	0.97
QT interval, ms (IQR)	360 (100)	380 (110)	Unit = 20	1.0 (0.9–1.1)	0.17
QTc interval, ms (IQR)	430 (68)	452 (88)	Unit = 20	1.1 (1.1–1.2)	<0.0001
QTc ≥500 ms	37 (12.7)	41 (30.8)	Yes vs. No	2.2 (1.4–3.4)	0.0002
Diagnosis of LQT3, n (%)	70 (23.1)	57 (41.0)	Yes vs. No	1.8 (1.2–2.7)	0.001
Diagnosis of sinus node dysfunction, n (%)	12 (4.0)	11 (7.9)	Yes vs. No	1.5 (0.7–3.1)	0.18
Diagnosis of AV block (any grade), n (%)	93 (30.8)	59 (42.4)	Yes vs. No	1.7 (1.2–2.6)	0.003
Diagnosis of RBBB (any grade), n (%)	122 (40.4)	66 (47.5)	Yes vs. No	1.5 (1.0–2.1)	0.03
Diagnosis of LBBB (any grade), n (%)	9 (3.0)	8 (5.8)	Yes vs. No	2.2 (0.9–4.9)	0.05
Diagnosis of SVT, n (%)	4 (1.3)	11 (7.9)	Yes vs. No	4 (1.9–8.9)	0.0002
Diagnosis of spontaneous BrS1, n (%)	24 (7.9)	14 (10.1)	Yes vs. No	1.2 (0.7–2.3)	0.42
Genetic characteristics					
Genotype					
Single SCN5A mutation, n (%)	299 (98.7)	131 (94.2)	Reference	1	0.004
Double SCN5A mutation, n (%)	1 (0.3)	2 (1.4)	vs. single	10.3 (1.8–58.7)	
Compound mutation, n (%)	3 (1.0)	6 (4.3)	vs. single	2.2 (0.8–6.2)	
Mutation type					
Non-missense pathogenic mutation, n (%)	74 (24.4)	39 (28.1)	Reference	1	0.52
Missense pathogenic mutation, n (%)	200 (66.0)	83 (59.7)	vs. non-missense	0.84 (0.54–1.31)	
Unknown functional effect, n (%)	29 (9.6)	17 (12.2)	vs. non-missense	1.03 (0.53–2.00)	
Mutation location (domains)					
N-terminus location, n (%)	4 (1.3)	3 (2.2)	vs. DI domain	1.3 (0.3–5.6)	<0.0001
DI domain, n (%)	37 (12.2)	27 (19.4)	Reference	1	
DI/DII interdomain linker, n (%)	18 (5.9)	8 (5.8)	vs. DI domain	0.7 (0.3–1.9)	
DII domain, n (%)	29 (9.6)	9 (6.5)	vs. DI domain	0.5 (0.2–1.1)	
DII/DIII interdomain linker, n (%)	22 (7.3)	8 (5.8)	vs. DI domain	0.5 (0.2–1.2)	
DIII domain, n (%)	49 (16.2)	19 (13.7)	vs. DI domain	0.5 (0.2–1.0)	
DIII/DIV interdomain linker, n (%)	15 (5.0)	13 (9.4)	vs. DI domain	1.3 (0.5–3.2)	
DIV domain, n (%)	40 (13.2)	31 (22.3)	vs. DI domain	1.4 (0.7–2.8)	
C-terminus, n (%)	89 (29.4)	21 (15.1)	vs. DI domain	0.3 (0.1–0.5)	
Mutation location (segments, n = 241)					
S1–S4, n (%)	51 (32.9)	29 (33.7)	Reference	1	0.52
S5–S6, n (%)	104 (67.1)	57 (66.3)	vs. S1–S4	1.1 (0.7–1.9)	
Mutation functional effect					
Loss of function, n (%)	126 (41.6)	52 (37.4)	Reference	1	<0.0001
Gain of function, n (%)	46 (15.2)	41 (29.5)	vs. loss-of-function	2.3 (1.4–3.9)	

Continued

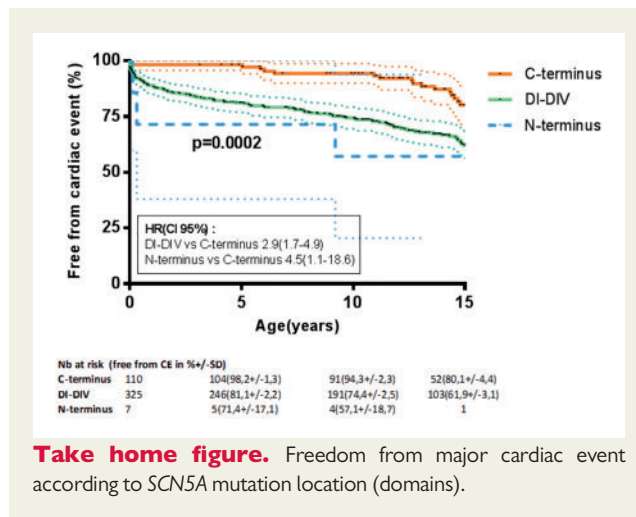
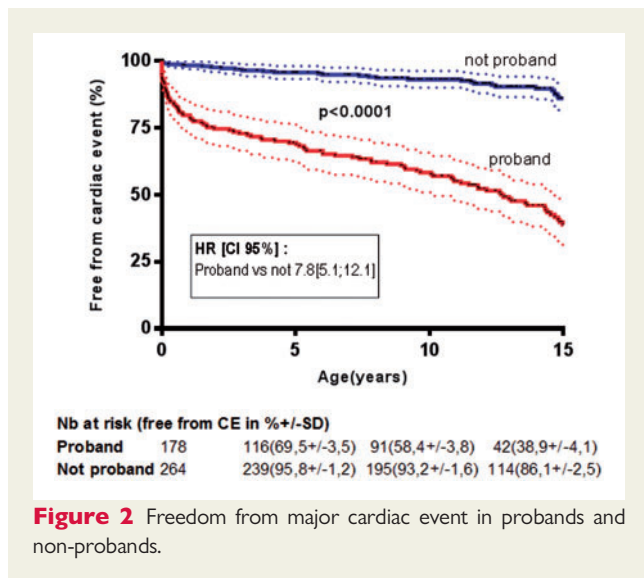
Table 1 Continued

	No MCE (n = 303)	MCE (n = 139)	Analysis	HR (95% CI)	P-value
Gain and loss, n (%)	71 (23.4)	14 (10.1)	vs. loss-of-function	0.4 (0.2–0.8)	
Unknown functional effect, n (%)	60 (19.8)	32 (23.0)	vs. loss-of-function	1.2 (0.7–2.1)	

AV block, atrioventricular block; BrS-1, Brugada syndrome type 1; CE, cardiac event; DCM, dilated cardiomyopathy; FH, family history; FU, follow-up; ICD, implantable cardioverter-defibrillator; LBBB, left bundle branch block; LQT3, long QT syndrome type 3; PCCD, progressive cardiac conduction defect; PM, pacemaker; QTc, corrected QT value; RBBB, right bundle branch block; SCD, sudden cardiac death; SSS, sick sinus syndrome; SVT, supraventricular tachycardia.

^aCox model is not applicable when subgroups contain no event. In this later case, we presented log-rank test.

Bold values are statistically significant P-values.



mutation, and 92 (21%) had a VUS. Although VUS patients were more frequently probands ($P=0.003$), their clinical characteristics did not differ from those of patients with a variant of known functional effect (Supplementary material online, Table S6). Most variants were missense pathogenic mutations (64%), whereas 25% were non-missense pathogenic mutations (truncation mutations: 18%, in-frame mutations: 7%). Topological location of mutations is shown in Supplementary material online, Figure S2.

Genotype–phenotype correlations

Mutation topological location

Patients with a mutation in the C-terminus domain ($n=110$) were less frequently probands ($P=0.03$), were diagnosed later in life ($P=0.01$), were less frequently symptomatic at diagnosis ($P=0.001$), had less MCEs ($P=0.0002$) and less appropriate ICD shocks ($P=0.03$) during follow-up (Supplementary material online, Table S7 and Take home Figure). No significant difference was found when comparing variants localized in S1–S4 to those localized in S5–S6 in the relevant 241 patients (Supplementary material online, Table S8).

Mutation functional effect

Children with a gain-of-function *SCN5A* mutation mainly presented with a baseline negative ECG phenotype (45%) or isolated LQT3 (26%); those with a loss-of-function mutation presented mainly with

isolated PCCD (38%), negative ECG phenotype (27%), or overlap phenotype (19%) at baseline; and those with a both gain- and loss-of-function mainly had negative ECG phenotype (35%), isolated PCCD (22%), isolated LQT3 (12%), or overlap phenotype (14%) (Supplementary material online, Table S9). Comparison between groups by looking at the functional effect of the mutation (gain of function, loss of function, or both) demonstrated that gain-of-function mutation carriers were more likely to have a cardiac arrest as first presentation ($P<0.001$) and a greater rate of both MCEs during follow-up ($P<0.001$) and ICD implantation ($P<0.001$).

Mutation type

Non-missense mutation were more frequently identified in case of isolated PCCD ($P<0.006$) but less frequently found in case of negative ECG phenotype ($P<0.007$) (Supplementary material online, Table S10). The following clinical parameters differed according to mutation type: age at diagnosis ($P=0.02$), proportion of diagnosis ≤ 1 year ($P=0.02$), FH of SCD/ICD ($P=0.03$), and FH of PCCD/PM ($P=0.001$), as did the following baseline phenotypes: isolated PCCD ($P=0.006$) and negative ECG phenotype ($P=0.007$) (Supplementary material online, Table S10). However, the type of mutation did not change the risk of MCE during follow-up.

Univariate risk analysis

The risk of MCE during follow-up was related to phenotype (Table 1). Age ≤ 1 year at diagnosis [HR (95% CI) 11.3 (6.7–18.9), $P<0.0001$],

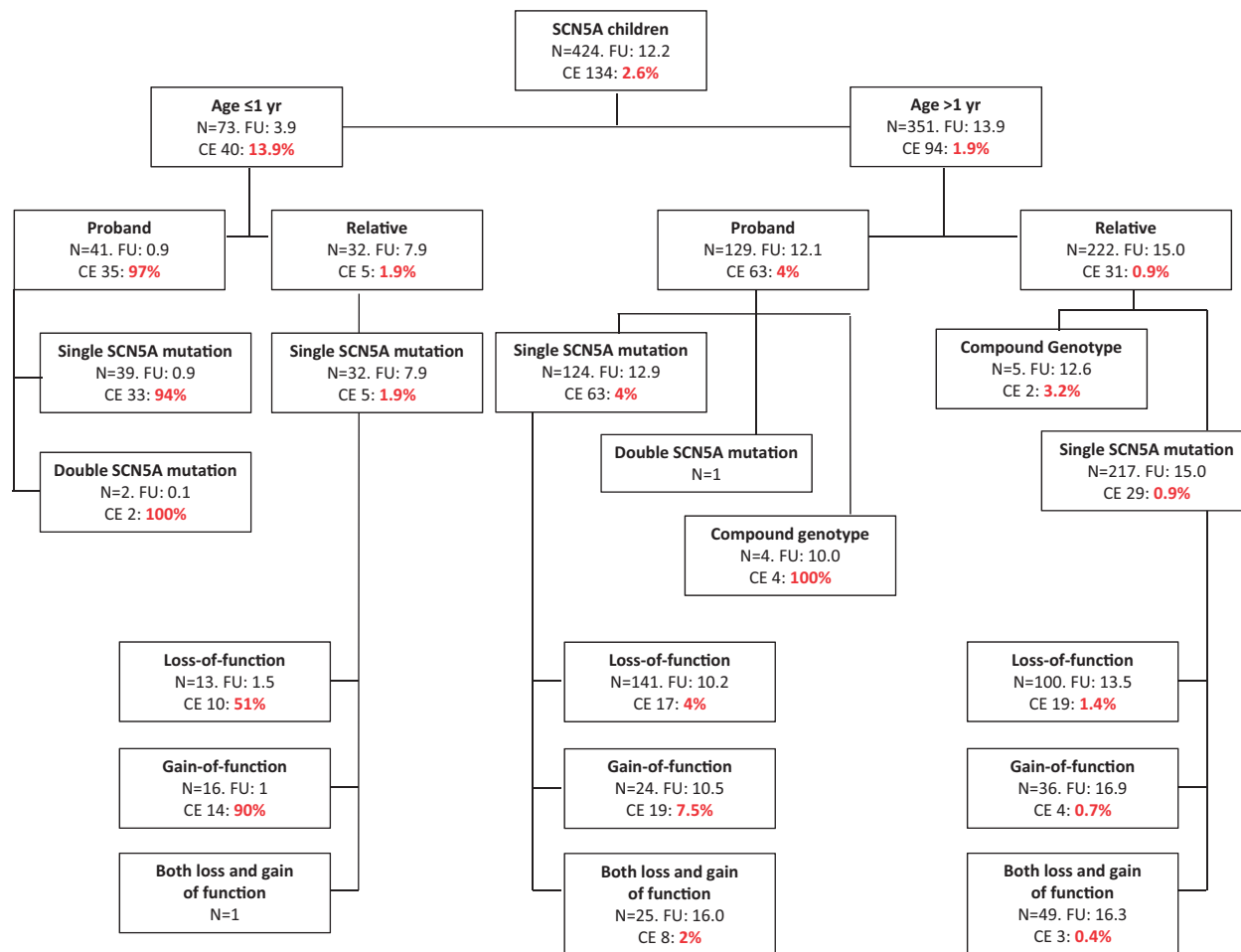


Figure 3 Mean event rate per year according to risk factors identified on multivariate analysis. FU, follow-up, %, mean event rate per year.

proband status [HR (95% CI) 7.8 (5.1–12.1), $P < 0.0001$] (Figure 2), supraventricular tachycardia [HR (95% CI) 4.0(1.9–8.9), $P = 0.0002$], baseline QTc ≥ 500 ms [HR (95% CI) 2.2(1.4–3.4), $P = 0.0002$], and AVB of any type [HR (95% CI) 1.7(1.2–2.6), $P = 0.003$] were predictors of MCEs. The effect of baseline ECG phenotype on the occurrence of MCE varied with age and the assumption of proportional hazards was not respected.

Occurrence of MCE also differed according to genotype ($P = 0.004$) [double vs single mutation: HR (95% CI) 10.3 (1.8–58.7); compound vs. single mutation: HR (95% CI) 2.2 (0.8–6.2)] (Table 1), gain-of-function mutation [HR (95% CI) 2.3 (1.4–3.9), $P < 0.0001$] and C-terminus mutation location [HR (95% CI) 0.3 (0.1–0.5), $P < 0.0001$] (Supplementary material online, Figure S3). Mutation type did not associate with outcomes ($P = 0.52$) (Supplementary material online, Figure S4).

Five SCN5A mutations correlated with specific clinical characteristics (Supplementary material online, Table S11). For instance, p.Glu1784Lys was associated with a lower risk of CE [$P = 0.0002$, HR (95% CI) 3.7 (1.8–7.6)], whereas the presence of p.Val411Met or p.Val1763Met was associated with a higher risk of CE [$P < 0.0001$, HR (95% CI) 5.1 (2.3–11.4) and $P < 0.0001$, HR (95% CI): 15.4 (5.4–43.4), respectively].

Multivariable analysis

A multivariable analysis stratified by baseline phenotype and adjusted on age ≤ 1 year at diagnosis and proband status (interaction, $P = 0.0002$), genotype ($P = 0.03$), and mutation functional effect ($P = 0.001$), showed that age ≤ 1 year at diagnosis in probands [$P < 0.0001$; HR (95% CI) 35.4 (16.2–77.6)], compound mutation [$P = 0.03$; HR (95% CI) 3.7 (1.2–12.0)], age ≤ 1 year at diagnosis in non-probands [$P = 0.03$; HR (95% CI) 3.2 (1.1–9.1)] and mutation with both gain- and loss-of-function [$P = 0.04$; HR (95% CI) 0.5 (0.2–0.9)] were independent risk factors for first CE (Supplementary material online, Table S12). Quantifiable indication of risk of events in an SCN5A mutation-positive child is presented in Supplementary material online, Figure S5.

Discussion

This study reports the clinical evaluation and follow-up of the largest paediatric population of SCN5A-mutation-positive individuals reported to date. We presented a highly symptomatic cohort with

SCD and ACA in 14%, syncope in 16%, and events during follow-up in 31%. Cardiac conduction disorder was the most prevalent phenotype. Age ≤ 1 year at diagnosis in probands, compound genotype, age ≤ 1 year at diagnosis in non-probands, and both gain- and loss-of-function *SCN5A* mutation were independent predictors of MCE. We also found that asymptomatic negative ECG phenotype children have a good prognosis, whereas previously symptomatic children with a negative ECG phenotype may undergo recurrent events even under treatment.

Clinical characteristics

The risk for life-threatening arrhythmias was higher in previously symptomatic patients, as previously shown in young BrS^{19,20} and LQT3 patients.^{21,22} We found no gender difference, in phenotype or in the risk for a MCE. Unlike previous adult studies where BrS was predominant in male subjects²³ and life-threatening events were higher among LQT3 men,²⁴ our results are concordant with previous smaller paediatric reports^{19,25,26} and the contradiction might be explained by similarities in sex hormones between prepubertal boys and girls. However, the underlying molecular mechanisms are still poorly understood.²⁷

In our series, more than one-third of isolated PCCD patients experienced MCE, the first of which being cardiac arrest in a high proportion of cases. Phenotypic expression of *SCN5A* mutations may vary from individual to individual and has an age-dependent onset.²⁸ Although there is no genotype-based risk stratification for PCCD patients, the occurrence of tachyarrhythmia and SCD was expected to be more frequent in case of loss-of-function *SCN5A* mutation, as per *SCN5A*-associated BrS that is a similar disease entity.²⁹ This was also suggested by familial reports of overlapping phenotypes of BrS1, LQTS, and PCCD^{3,12} and the observation that BrS patients with *SCN5A* mutations exhibit more conduction abnormalities and have a higher risk for MCEs.³⁰ Our results demonstrate that some isolated PCCD patients are at increased risk of SCD indeed, even at an early age and even if an isolated PCCD phenotype is maintained throughout follow-up, an AVB of any type being an univariate risk factor for CE. Children diagnosed with an AVB of any type should therefore be offered genetic screening; when a *SCN5A* mutation is diagnosed, ICD therapy should be discussed in this high-risk group in case of additional risk factors that are age ≤ 1 year at diagnosis in probands, compound mutation, age ≤ 1 year at diagnosis in non-probands and *SCN5A* mutation with both gain- and loss-of-function.

There is also limited data on *SCN5A* genotype positive children with a negative ECG phenotype.^{12,14} We found that the vast majority of those who are asymptomatic at diagnosis have a good long-term prognosis; however, they need to be followed, as negative ECG phenotype patients may develop a phenotype over time. Negative ECG phenotype children can also present with symptoms; close follow-up and ICD implantation should be considered in symptomatic *SCN5A* mutation-positive children, even if displaying a negative ECG phenotype, because a substantial proportion of them will experience further recurrent events, even under appropriate treatment.

Correlation between genotype and phenotype

Unlike a previous small report of loss-of-function cardiac sodium channelopathies that indicated that missense pathogenic variants were more common,²⁵ non-missense pathogenic variants were over-represented in isolated PCCD in our much larger sample. This is concordant with the role of haploinsufficiency in causing greater impairment of I_{Na} and more severe phenotype leading to PCCD. Phenotype correlation of *SCN5A* mutation-positive subjects, based on variant location has not been possible before due to small numbers.³¹ We found that the N-terminus domain, the DI-DIV region and the C-terminus domain were not over-represented amongst the five main ECG phenotypes. No difference appeared when considering the six segments of the transmembrane domains. However, in a recent case/control study, Kapplinger et al.¹⁷ were able to identify regions of Nav1.5 associated with a high probability of pathogenicity in both BrS and LQT3. In their study, the transmembrane region yielded an overrepresentation of BrS-associated variants, whereas the DIII/DIV interdomain linker and the S3-S5 + 6 segment of all transmembrane domains hosted an over-representation of LQT3-associated variants.¹⁷ These differences are likely due to ascertainment biases inherent to each study design.

Clinical severity: clinical and genetic predictors

The high incidence of MCEs in our cohort was concordant with a previous small LQT3 paediatric multicentre international study²¹ and a recent multicentre series of 391 adult and paediatric LQT3 patients.²² However, the burden of events was higher than reported by other LQT3 or BrS series in the past.^{26,32,33} The rate of SCD or ACA in our cohort was 14%, similar to other recent reports on LQT3 patients^{21,22} but significantly higher than that reported in BrS children.^{19,26,34} This may reflect an over-representation of LQT3 phenotypes in our cohort, as LQT3 patients who experience MCE during the first year of life are at high risk for subsequent MCEs.^{32,35,36} Indeed, we found that ACA was the first symptom in 23% of the 47 isolated LQT3 children who exhibited a 7% annual rate of CE per year throughout follow-up, although only 1 (4%) was on beta-blocker at the time of the first MCE. Moreover, the two *SCN5A* mutations associated with an increased risk of MCEs in our series, namely p.Val411Met and p.Val1763Met were both gain-of-function mutations.

SCN5A mutations localizing to the transmembrane regions or the N-terminus were associated with a higher risk for CE compared to the C-terminus. This is an important finding that may help geneticists and physicians counselling young affected individuals and their families.

It is recognized that double *SCN5A* mutation carriers have a more severe phenotype with longer QTc intervals, a younger age at diagnosis, and more CEs despite therapy.³³

Schwartz et al.³⁶ first raised the issue of different response of LQT3 patients to beta-blockers and/or LCSD between infants with MCEs in the first year of life and those presenting later. This concept was then confirmed by data from the International LQTS Registry showing that patients with an ACA during their first year of life had a very high risk for subsequent ACA or SCD during their next 10 years

of life and that beta-blockers might not be effective in preventing fatal MCEs in this high-risk subset.³⁷ Our results extend this observation to all paediatric *SCN5A* genotype positive subjects, whatever their ECG phenotype, as we found that both age ≤ 1 year at diagnosis in probands and age ≤ 1 year at diagnosis in non-probands were independent risk factors for first CE. A significant subset of these patients might represent *de novo* mutations, which are usually associated with greater physico-chemical difference and are more likely to be more severe in effect than inherited mutations.³⁸ This is in keeping with the observation of *de novo* mutations in the *SCN5A* gene associated with early onset of sudden infant death.^{9,10,39} Our observation may therefore be due to a clustering of *de novo* mutations⁴⁰ and *SCN5A* mutation-positive patients with no FH constitute a subgroup at high-risk of ACA and arrhythmic events and should be treated accordingly.

Conclusions

In this large paediatric cohort of *SCN5A* genotype positive patients, cardiac conduction disorders were the most prevalent phenotype. Symptomatic individuals and LQT3 patients had the worst prognosis. Age ≤ 1 year at diagnosis in probands was associated with the highest risk. However, both negative ECG phenotype children and isolated PCCD children can also present with symptoms and these patients need to be accurately treated and followed. Compound genotype with associated mutation in another gene and for the first time variant topological location were independent risk factors for CEs. These findings offer therapeutic opportunity for determining risk in these vulnerable young patients.

Supplementary material

Supplementary material is available at *European Heart Journal* online.

Acknowledgements

The authors gratefully acknowledge the European Reference Network on Rare and Complex Diseases of the Heart (ERN GUARD-HEART).

Funding

French Society of Cardiology (to A.-E.B.) a research grant the Foundation Bettencourt-Schueller (to A.-E.B.) and by research funds from Cardiac Risk in the Young (to A.-E.B., L.W., and E.R.B.); by the National Institute for Health Research Biomedical Research Centre at Great Ormond Street Hospital for Children NHS Foundation Trust and University College London (to J.P.K.); by the Mayo Clinic Windland Smith Rice Comprehensive Sudden Cardiac Death Program (to M.J.A.); by the Netherlands CardioVascular Research Initiative: the Dutch Heart Foundation, Dutch Federation of University Medical Centres, the Netherlands Organisation for Health Research and Development, and the Royal Netherlands Academy of Sciences (CVON Predict to A.A.W.).

Conflict of interest: M.J.A. is consultant for Audentes Therapeutics, Boston Scientific, Gilead Sciences, Invitae, Medtronic, MyoKardia, and St. Jude Medical. M.J.A. and Mayo Clinic have an equity/royalty relationship with AliveCor, Blue Ox Health Corporation, and Stemonix. However, none of these entities were involved in this study in any manner. None of

the authors has any financial relationships relevant to this article to disclose.

References

- Wang Q, Shen J, Splawski I, Atkinson D, Li Z, Robinson JL, Moss AJ, Towbin JA, Keating MT. *SCN5A* mutations associated with an inherited cardiac arrhythmia, long QT syndrome. *Cell* 1995;**80**:805–811.
- Chen Q, Kirsch GE, Zhang D, Brugada R, Brugada J, Brugada P, Potenza D, Moya A, Borggrefe M, Breithardt G, Ortiz-Lopez R, Wang Z, Antzelevitch C, O'Brien RE, Schulze-Bahr E, Keating MT, Towbin JA, Wang Q. Genetic basis and molecular mechanism for idiopathic ventricular fibrillation. *Nature* 1998;**392**:293–296.
- Kyndt F, Probst V, Potet F, Demolombe S, Chevallier JC, Baro I, Moisan JP, Boisseau P, Schott JJ, Escande D, Le Marec H. Novel *SCN5A* mutation leading either to isolated cardiac conduction defect or Brugada syndrome in a large French family. *Circulation* 2001;**104**:3081–3086.
- Schott JJ, Alshinawi C, Kyndt F, Probst V, Hoorntje TM, Hulsbeek M, Wilde AA, Escande D, Mannens MM, Le Marec H. Cardiac conduction defects associate with mutations in *SCN5A*. *Nat Genet* 1999;**23**:20–21.
- Benson DW, Wang DW, Dymont M, Knilans TK, Fish FA, Strieper MJ, Rhodes TH, George AL. Congenital sick sinus syndrome caused by recessive mutations in the cardiac sodium channel gene (*SCN5A*). *J Clin Invest* 2003;**112**:1019–1028.
- Olson TM, Michels VV, Ballew JD, Reyna SP, Karst ML, Herron KJ, Horton SC, Rodeheffer RJ, Anderson JL. Sodium channel mutations and susceptibility to heart failure and atrial fibrillation. *JAMA* 2005;**293**:447–454.
- Laurent G, Saal S, Amarouch MY, Béziau DM, Marsman RFJ, Faivre L, Barc J, Dina C, Bertaux G, Barthez O, Thauvin-Robinet C, Charron P, Fressart V, Maltret A, Villain E, Baron E, Mérot J, Turpault R, Coudière Y, Charpentier F, Schott JJ, Loussouarn G, Wilde AAM, Wolf J-E, Baró I, Kyndt F, Probst V. Multifocal ectopic Purkinje-related premature contractions: a new *SCN5A*-related cardiac channelopathy. *J Am Coll Cardiol* 2012;**60**:144–156.
- McNair WP, Ku L, Taylor MRG, Fain PR, Dao D, Wolfel E, Mestroni L; Familial Cardiomyopathy Registry Research Group. *SCN5A* mutation associated with dilated cardiomyopathy, conduction disorder, and arrhythmia. *Circulation* 2004;**110**:2163–2167.
- Schwartz PJ, Priori SG, Dumaine R, Napolitano C, Antzelevitch C, Stramba-Badiale M, Richard TA, Berti MR, Bloise R. A molecular link between the sudden infant death syndrome and the long-QT syndrome. *N Engl J Med* 2000;**343**:262–267.
- Ackerman MJ, Siu BL, Sturner WQ, Tester DJ, Valdivia CR, Makielski JC, Towbin JA. Postmortem molecular analysis of *SCN5A* defects in sudden infant death syndrome. *JAMA* 2001;**286**:2264–2269.
- Wong LCH, Behr ER. Sudden unexplained death in infants and children: the role of undiagnosed inherited cardiac conditions. *Europace* 2014;**16**:1706–1713.
- Priori SG, Wilde AA, Horie M, Cho Y, Behr ER, Berul C, Blom N, Brugada J, Chiang C-E, Huikuri H, Kannankeril P, Krahn A, Leenhardt A, Moss A, Schwartz PJ, Shimizu W, Tomaselli G, Tracy C. HRS/EHRA/APHRS expert consensus statement on the diagnosis and management of patients with inherited primary arrhythmia syndromes: document endorsed by HRS, EHRA, and APHRS in May 2013 and by ACCF, AHA, PACES, and AEP in June 2013. *Heart Rhythm* 2013;**10**:1932–1963.
- Kato K, Makiyama T, Wu J, Ding W-G, Kimura H, Naiki N, Ohno S, Itoh H, Nakanishi T, Matsuura H, Horie M. Cardiac channelopathies associated with infantile fatal ventricular arrhythmias: from the cradle to the bench. *J Cardiovasc Electrophysiol* 2014;**25**:66–73.
- Harris BU, Miyake CY, Motonaga KS, Dubin AM. Diagnosis and management of pediatric brugada syndrome: a survey of pediatric electrophysiologists. *Pacing Clin Electrophysiol PACE* 2014;**37**:638–642.
- Wang Q, Li Z, Shen J, Keating MT. Genomic organization of the human *SCN5A* gene encoding the cardiac sodium channel. *Genomics* 1996;**34**:9–16.
- Meregalli PG, Tan HL, Probst V, Koopmann TT, Tanck MW, Bhuiyan ZA, Sacher F, Kyndt F, Schott JJ, Albuissin J, Mabo P, Bezzina CR, Le Marec H, Wilde AAM. Type of *SCN5A* mutation determines clinical severity and degree of conduction slowing in loss-of-function sodium channelopathies. *Heart Rhythm* 2009;**6**:341–348.
- Kapplinger JD, Giudicessi JR, Ye D, Tester DJ, Callis TE, Valdivia CR, Makielski JC, Wilde AA, Ackerman MJ. Enhanced classification of Brugada syndrome-associated and long-QT syndrome-associated genetic variants in the *SCN5A*-encoded Na(v)1.5 cardiac sodium channel. *Circ Cardiovasc Genet* 2015;**8**:582–595.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehms HL; ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015;**17**:405–424.

19. Andorin A, Behr ER, Denjoy I, Crotti L, Dagradi F, Jesel L, Sacher F, Petit B, Mabo P, Maltret A, Wong LCH, Degand B, Bertaux G, Maury P, Dulac Y, Delasalle B, Gourraud J-B, Babuty D, Blom NA, Schwartz PJ, Wilde AA, Probst V. Impact of clinical and genetic findings on the management of young patients with Brugada syndrome. *Heart Rhythm* 2016;**13**:1274–1282.
20. Gonzalez Corcia MC, Sieira J, Sarkozy A, Asmundis C, de Chierchia G-B, Hernandez Ojeda J, Pappaert G, Brugada P. Brugada syndrome in the young: an assessment of risk factors predicting future events. *Europace* 2017;**19**:1864–1873.
21. Blaufox AD, Tristani-Firouzi M, Seslar S, Sanatani S, Trivedi B, Fischbach P, Paul T, Young M-L, Tisma-Dupanovic S, Silva J, Cuneo B, Fournier A, Singh H, Tanel RE, Etheridge SP. Congenital long QT 3 in the pediatric population. *Am J Cardiol* 2012;**109**:1459–1465.
22. Wilde AAM, Moss AJ, Kaufman ES, Shimizu W, Peterson DR, Benhorin J, Lopes C, Towbin JA, Spazzolini C, Crotti L, Zareba W, Goldenberg I, Kanter J, Robinson JL, Qi M, Hofman N, Tester DJ, Bezzina CR, Alders M, Aiba T, Kamakura S, Miyamoto Y, Andrews ML, McNitt S, Polonsky B, Schwartz PJ, Ackerman MJ. Clinical aspects of type 3 long-QT syndrome: an International Multicenter Study. *Circulation* 2016;**134**:872–882.
23. Eckardt L, Probst V, Smits JPP, Bahr ES, Wolpert C, Schimpf R, Wichter T, Boisseau P, Heinecke A, Breithardt G, Borggrefe M, LeMarec H, Böcker D, Wilde AAM. Long-term prognosis of individuals with right precordial ST-segment-elevation Brugada syndrome. *Circulation* 2005;**111**:257–263.
24. Priori SG, Schwartz PJ, Napolitano C, Bloise R, Ronchetti E, Grillo M, Vicentini A, Spazzolini C, Nastoli J, Bottelli G, Folli R, Cappelletti D. Risk stratification in the long-QT syndrome. *N Engl J Med* 2003;**348**:1866–1874.
25. Chockalingam P, Clur S-AB, Breur JMP, Kriebel T, Paul T, Rammeloo LA, Wilde AAM, Blom NA. The diagnostic and therapeutic aspects of loss-of-function cardiac sodium channelopathies in children. *Heart Rhythm* 2012;**9**:1986–1992.
26. Probst V, Denjoy I, Merregalli PG, Amirault J-C, Sacher F, Mansourati J, Babuty D, Villain E, Victor J, Schott J-J, Lupoglazoff J-M, Mabo P, Veltmann C, Jesel L, Chevalier P, Clur S-AB, Haissaguerre M, Wolpert C, Le Marec H, Wilde AAM. Clinical aspects and prognosis of Brugada syndrome in children. *Circulation* 2007;**115**:2042–2048.
27. Shimizu W, Matsuo K, Kokubo Y, Satomi K, Kurita T, Noda T, Nagaya N, Suyama K, Aihara N, Kamakura S, Inamoto N, Akahoshi M, Tomoike H. Sex hormone and gender difference—role of testosterone on male predominance in Brugada syndrome. *J Cardiovasc Electrophysiol* 2007;**18**:415–421.
28. Ackerman MJ, Priori SG, Willems S, Berul C, Brugada R, Calkins H, Camm AJ, Ellinor PT, Gollob M, Hamilton R, Hershberger RE, Judge DP, Le Marec H, McKenna WJ, Schulze-Bahr E, Semsarian C, Towbin JA, Watkins H, Wilde A, Wolpert C, Zipes DP. HRS/EHRA expert consensus statement on the state of genetic testing for the channelopathies and cardiomyopathies this document was developed as a partnership between the Heart Rhythm Society (HRS) and the European Heart Rhythm Association (EHRA). *Heart Rhythm* 2011;**8**:1308–1339.
29. Gray B, Behr ER. New insights into the genetic basis of inherited arrhythmia syndromes. *Circ Cardiovasc Genet* 2016;**9**:569–577.
30. Yamagata K, Horie M, Aiba T, Ogawa S, Aizawa Y, Ohe T, Yamagishi M, Makita N, Sakurada H, Tanaka T, Shimizu A, Hagiwara N, Kishi R, Nakano Y, Takagi M, Miyama T, Ohno S, Fukuda K, Watanabe H, Morita H, Hayashi K, Kusano K, Kamakura S, Yasuda S, Ogawa H, Miyamoto Y, Kapplinger JD, Ackerman MJ, Shimizu W. Genotype-phenotype correlation of SCN5A mutation for the clinical and electrocardiographic characteristics of probands with Brugada syndrome: a Japanese Multicenter Registry. *Circulation* 2017;**135**:2255–2270.
31. Schwartz PJ, Dagradi F. Management of survivors of cardiac arrest—the importance of genetic investigation. *Nat Rev Cardiol* 2016;**13**:560–566.
32. Priori SG, Napolitano C, Schwartz PJ, Grillo M, Bloise R, Ronchetti E, Moncalvo C, Tulipani C, Veia A, Bottelli G, Nastoli J. Association of long QT syndrome loci and cardiac events among patients treated with beta-blockers. *Jama* 2004;**292**:1341–1344.
33. Schwartz PJ, Spazzolini C, Priori SG, Crotti L, Vicentini A, Landolina M, Gasparini M, Wilde AAM, Knops RE, Denjoy I, Toivonen L, Mönnig G, Al-Fayyadh M, Jordaens L, Borggrefe M, Holmgren C, Brugada P, De Roy L, Hohnloser SH, Brink PA. Who are the long-QT syndrome patients who receive an implantable cardioverter-defibrillator and what happens to them? Data from the European Long-QT Syndrome Implantable Cardioverter-Defibrillator (LQTS ICD) Registry. *Circulation* 2010;**122**:1272–1282.
34. Conte G, Dewals W, Sieira J, Asmundis C, de Ciconte G, Chierchia G-B, Di Giovanni G, Baltogiannis G, Saitoh Y, Levinstein M, La Meir M, Wellens F, Pappaert G, Brugada P. Drug-induced brugada syndrome in children: clinical features, device-based management, and long-term follow-up. *J Am Coll Cardiol* 2014;**63**:2272–2279.
35. Schwartz PJ, Priori SG, Spazzolini C, Moss AJ, Vincent GM, Napolitano C, Denjoy I, Guicheney P, Breithardt G, Keating MT, Towbin JA, Beggs AH, Brink P, Wilde AA, Toivonen L, Zareba W, Robinson JL, Timothy KW, Corfield V, Wattanasirichaigoon D, Corbett C, Haverkamp W, Schulze-Bahr E, Lehmann MH, Schwartz K, Coumel P, Bloise R. Genotype-phenotype correlation in the long-QT syndrome: gene-specific triggers for life-threatening arrhythmias. *Circulation* 2001;**103**:89–95.
36. Schwartz PJ, Spazzolini C, Crotti L. All LQT3 patients need an ICD: true or false? *Heart Rhythm* 2009;**6**:113–120.
37. Spazzolini C, Mullally J, Moss AJ, Schwartz PJ, McNitt S, Ouellet G, Fugate T, Goldenberg I, Jons C, Zareba W, Robinson JL, Ackerman MJ, Benhorin J, Crotti L, Kaufman ES, Locati EH, Qi M, Napolitano C, Priori SG, Towbin JA, Vincent GM. Clinical implications for patients with long QT syndrome who experience a cardiac event during infancy. *J Am Coll Cardiol* 2009;**54**:832–837.
38. Brunklaus A, Ellis R, Reavey E, Semsarian C, Zuberi SM. Genotype phenotype associations across the voltage-gated sodium channel family. *J Med Genet* 2014;**51**:650–658.
39. Wedekind H, Smits JP, Schulze-Bahr E, Arnold R, Veldkamp MW, Bajanowski T, Borggrefe M, Brinkmann B, Warnecke I, Funke H, Bhuiyan ZA, Wilde AA, Breithardt G, Haverkamp W. De novo mutation in the SCN5A gene associated with early onset of sudden infant death. *Circulation* 2001;**104**:1158–1164.
40. Francioli LC, Polak PP, Koren A, Menelaou A, Chun S, Renkens I, van Duijn CM, Swertz M, Wijmenga C, van Ommen G, Slagboom PE, Boomsma DI, Ye K, Guryev V, Arndt PF, Kloosterman WP, de Bakker PIW, Sunyaev SR. Genome-wide patterns and properties of de novo mutations in humans. *Nat Genet* 2015;**47**:822–826.